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Cladosporium spp. (Cladosporiaceae) isolated from *Eucommia ulmoides* in China

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Abstract

Eucommia ulmoides is a rare tree species in China with high medicinal and gum value. Nine strains of hyphomycetous fungi were isolated from the leaf litter of *E. ulmoides* in Guizhou Province. Preliminary identifications based on ITS indicated that they belong to the genus *Cladosporium*. Morphology and phylogenetic analyses based on the internal transcribed spacer regions (ITS) of the nrDNA, the partial translation elongation factor 1-α (*tef1*) gene and partial of actin (*act*) gene confirmed that the strains represent four species, including two novel taxa, viz., *Cladosporium eucommiae* and *C. guizhouense* and two new substrate records for known species.

Keywords

Asexual morphs, new species, phylogeny, taxonomy

Introduction

Eucommia ulmoides Oliver ('du-zhong' in Chinese), the single extant species of *Eucommiaceae* (related to *Ulmaceae*), is a dioecious, wind-pollinated tree evenly distributed in mixed mesophytic forest habitats of valleys, hills, and low mountains in central and eastern China (Cronquist 1981; Zhang 2016). *E. ulmoides* is widely cultivated in China and other countries owing to its high medicinal and gum value.

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The fungal genus *Cladosporium* was established by Link (1816). *Cladosporium* (*Cladosporiaceae*) is a ubiquitous genus in *Dothideomycetes* (Abdollahzadeh et al. 2020). This genus is widely distributed throughout the world and isolated from various sources such as air, soil, plants, food, debris, cloth, paint and other organic materials (Ellis 1977; Bensch et al. 2010, 2012, 2018; Temperini et al. 2018; Chung et al. 2019). Most *Cladosporium* species are saprobic (Bensch et al. 2010), and they occur on various senescing and dead leaves and stems of herbaceous and woody plants (Brown et al. 1998; El-Morsy 2000). The morphology of *Cladosporium* is mainly characterized by its asexual morph, which comprises differentiated conidiophores producing acropetal chains of conidia from mono- or polyblastic conidiogenous cells (Isabel et al. 2021). Both the conidiogenous cells and conidia show conidiogenous loci (scars) with a distinctive coronal structure, which is composed of a central convex dome surrounded by a raised periphery, usually thickened, refractive and dark (David 1997; Isabel et al. 2021). A molecular approach combined with morphological features has recognized more than 230 species in *Cladosporium*, which are grouped into three species complexes, i.e., the *C. cladosporioides*, *C. herbarum* and *C. sphaerospermum* complex (Schubert et al. 2007; Bensch et al. 2010, 2012, 2015, 2018; Sandoval-Denis et al. 2016; Marin-Felix et al. 2017).

In a recent research program, we have carried out a survey of micro-fungi associated with *E. ulmoides* in a forest in China. In this study, four *Cladosporium* taxa were isolated from fallen leaves of this plant species in Guizhou Province, including two new species, namely *C. eucommiae* and *C. guizhouense* spp. nov., which are introduced based on morphology and phylogenetic analyses. Newly generated molecular data, descriptions and illustrations of *C. tenuissimum* and *C. perangustum* are also provided herein.

Materials and methods

Sample collection and fungal strains isolation

Fallen leaves of *E. ulmoides* were collected in a forest plantation of Guizhou University, Guiyang, Guizhou Province, China, in January 2021. The samples were stored in envelopes and several topsoil samples from the forest were stored in self-sealing bags, then taken back to the laboratory and photographed. Before isolation, collected leaves samples were sprayed two to three times with 75% ethanol to disinfect the leaf surface. Pure cultures of the fungi were obtained by single spore isolation (Chomnunti et al. 2014). Fungi in the soil samples were isolated by the dilution plate method (Zhang et al. 2015). A small amount of soil (1 g) per sample was collected and added to 9 mL of sterile water in a 15 mL sterile glass test tube. It was manually mixed and then the suspension was diluted to a series of concentrations (10^{-1}, 10^{-2}, 10^{-3}, 10^{-4}, 10^{-5} and 10^{-6}), and 100 μL from each concentration was spread onto 90-mm-diam

Petri dishes containing Synthesis of low nutrient Agar (SNA), Potato Dextrose Agar (PDA), Malt Extract Agar (MEA) and Oatmeal Agar (OA) (Zhang et al. 2017). These SNA, PDA, MEA and OA plates were incubated at constant temperature (25 °C) in a controlled temperature light incubator. Holotype specimens of the new species were conserved in the Herbarium of the Department of Plant Pathology, Agricultural College, Guizhou University (**HGUP**). Ex-type cultures were conserved in the Culture Collection at the Department of Plant Pathology, Agriculture College, Guizhou University, P.R. China (**GUCC**).

Morphological description

Pure cultures were grown on SNA, PDA, MEA and OA media in a constant temperature incubator (25 °C). Culture characteristics were recorded and examined using a dissecting microscope (LEICA S9i, Germany). The morphological observations and measurements on SNA were made using a Zeiss Scope 5 compound microscope (Axioscope 5, China) with an attached camera AxioCam 208 color (ZEN 3.0) and measurements were made using ZEN 3.0. Taxonomic information for the two new taxa were deposited in MycoBank (www.mycobank.org).

DNA extraction, PCR amplification and sequencing

Fresh mycelia were scraped from the PDA plates with a sterilized scalpel. Genomic DNA was extracted using Fungal gDNA Kit (Biomiga #GD2416, San Diego, California, USA) in accordance with the manufacturer's instructions. PCR amplification was performed in a 25 μL reaction volume following Liang et al. (2018). Primer pairs ITS4/ITS5 (White et al. 1990), EF1-728F/EF1-986R (Carbone and Kohn 1999) and ACT-512F/ACT-783R (Carbone and Kohn 1999) were used for ITS, *tef1* and *act*, respectively. The amplification procedures were performed using the method described by Halo et al. (2019). Purification and sequencing of these three gene loci were carried out by the SinoGenoMax company (Beijing, China).

Phylogeny

Sequences used in this study (Table 1) were assembled based on the closest matches from the BLASTn search results (https://blast.ncbi.nlm.nih.gov/Blast.cgi) and previous publications (Sandoval-Denis et al. 2016; Bensch et al. 2018; Halo et al. 2019). Alignments were conducted with the online version of MAFFT v. 7.307 (Katoh and Standley 2016), checked visually and improved manually where necessary using BioEdit v. 7.1.3.0 (Hall 1999). SequenceMatrix v. 1.7.8 (Vaidya et al. 2011) was used to concatenate the aligned sequences of the different loci. Ambiguous areas were excluded from the analysis using AliView (Larsson 2014) and gaps were coded as missing data.

Table 1. Taxa used for molecular phylogenetic analyses and their GenBank accession numbers. Newly generated sequences are in bold. ([T]) = ex-holotype strain, ([ET]) = ex-epitype strain, ([NT]) = ex-neotype strain.

Species	Strain number	Host	Country	GenBank Accession number		
				ITS	*tef1*	*act*
Cladosporium angulosum	CBS 140692[T]	Man, bronchoalveolar lavage fluid	USA	LN834425	LN834521	LN834609
C. angulosum	CPC 11526	Acacia mangium	Thailand	HM148127	HM148371	HM148616
C. anthropophilum	CBS 140685[T]	Man, bronchoalveolar lavage fluid	USA	LN834437	LN834533	LN834621
C. anthropophilum	CBS 117483	–	USA	HM148007	HM148248	HM148494
C. anthropophilum	CPC 22272	Indoor air sample, ship	USA	MF574171	MF574173	MF574175
C. cladosporioides	CBS 101367	Soil	Brazil	HM148002	HM148243	HM148489
C. cladosporioides	CBS 112388	Air, indoor environment	Germany	HM148003	HM148244	HM148490
C. cladosporioides	CBS 113738	*Grape bud*	USA	HM148004	HM148245	HM148491
C. colocasiae	CBS 386.64T	*Colocasia esculenta*	Taiwan	HM148067	HM148310	HM148555
C. colocasiae	CBS 119542	*Colocasia esculenta*	Japan	HM148066	HM148309	HM148554
***C. eucommiae* sp. nov.**	**GUCC 401.1[T]**	**Fallen leaves of *Eucommia ulmoides***	**China**	**OL587465**	**OL504966**	**OL519775**
***C. eucommiae* sp. nov.**	**GUCC 401.9**	**Fallen leaves of Eucommia ulmoides**	**China**	**ON334729**	**–**	**ON383337**
***C. guizhouense* sp. nov.**	**GUCC 401.7[T]**	**Fallen leaves of *Eucommia ulmoides***	**China**	**OL579741**	**OL504965**	**OL519780**
***C. guizhouense* sp. nov.**	**GUCC 401.8**	**Fallen leaves of *Eucommia ulmoides***	**China**	**ON334728**	**ON383470**	**ON383338**
C. magnoliigena	MFLUCC 18-1559[T]	*Magnolia grandiflora*	China	MK347813	MK340864	–
C. magnoliigena	MFLUCC 18-1557	*Magnolia grandiflora*	China	MK347811	MK340862	–
C. oxysporum	CBS 125991[T]	Soil, near the terracotta army	China	HM148118	HM148362	HM148607
C. oxysporum	CBS 126351	Indoor air	Venezuela	HM148119	HM148363	HM148608
C. perangustum	**GUCC 401.6**	**Fallen leaves of *Eucommia ulmoides***	**China**	**OL579742**	**OL581726**	**OL519779**
C. perangustum	CBS 125996[T]	*Cussonia sp.*	South Africa	HM148121	HM148365	HM148610
C. perangustum	CPC 12216	*Morus rubra*	Germany	HM148135	HM148379	HM148624
C. perangustum	CPC 14247	*Magnolia sp.*	USA	HM148145	HM148389	HM148634
C. perangustum	CPC 13870	*Teratosphaeria fibrillosa*	South Africa	HM148142	HM148386	HM148631
C. perangustum	DTO 323-E4	Indoor air	China	MF473180	MF473602	MF474028
C. perangustum	CPC 22297	Indoor air sample	USA	MF473172	MF473595	MF474020
C. rectoides	CBS 125994[T]	*Vitis flexuosa*	South Korea	HM148193	HM148438	HM148683
C. tenuissimum	**GUCC 401.2**	**Fallen leaves of *Eucommia ulmoides***	**China**	**OL579746**	**OL504967**	**OL519776**
C. tenuissimum	**GUCC 401.3**	**Fallen leaves of *Eucommia ulmoides***	**China**	**OL579745**	**OL505077**	**–**
C. tenuissimum	**GUCC 401.4**	**Fallen leaves of *Eucommia ulmoides***	**China**	**OL579744**	**OL581724**	**OL519777**
C. tenuissimum	**GUCC 401.5**	**Fallen leaves of *Eucommia ulmoides***	**China**	**OL579743**	**OL581725**	**OL519778**
C. tenuissimum	CBS 125995[ET]	*Lagerstroemia sp.*	USA	HM148197	HM148442	HM148687
C. tenuissimum	CPC 12795	*Musa sp.*	Polynesia	HM148209	HM148454	HM148699
C. tenuissimum	CBS 126359	*Musa sp.*	USA	HM148198	HM148443	HM148688
C. tenuissimum	CPC 10882	*Gnaphalium affine*	South Korea	HM148204	HM148449	HM148694
C. tenuissimum	CPC 10538	*Musa sp.*	Mozambique	HM148202	HM148447	HM148692
C. tenuissimum	DTO 323-C5	Indoor air	China	MF473289	MF473712	MF474139
C. tenuissimum	CPC 13252	Rock	Australia	HM148216	HM148461	HM148706

Species	Strain number	Host	Country	GenBank Accession number		
				ITS	*tef1*	*act*
C. tenuissimum	CPC 13732	*Shorea siamensis*	Laos	HM148217	HM148462	HM148707
C. tenuissimum	CPC 14196	*Basella alba*, leaves	Laos	HM148218	HM148463	HM148708
C. xanthochromaticum	CPC 11609^{T}	Man, bronchoalveolar lavage fluid	USA	EF679356	EF679431	EF679508
C. xanthochromaticum	CBS 126364	*Erythrophleum chlorostachys*	Australia	HM148122	HM148366	HM148611
C. xylophilum	CBS 125997^{T}	*Picea abies*, dead wood	Russia	HM148230	HM148476	HM148721
C. langeronii	CBS 189.54^{T}	Man, mycosis	Brazil	DQ780379	JN906990	EF101357
C. neolangeronii	CBS 797.97^{T}	Indoor environment	Netherlands	MF473143	MF473576	MF473992

The Maximum Likelihood (ML) analyses were carried out at the CIPRES web portal (Miller et al. 2010) using RAxML (Stamatakis 2006). The tree search included 1,000 non-parametric bootstrap replicates and the best scoring tree was selected from suboptimal trees under the GTRGAMMA substitution model. The resulting replicates were plotted on to the best scoring tree obtained previously. Non-parametric bootstrap analysis was implemented with 1,000 duplicates. Maximum Parsimony (MP) analyses were performed with PAUP v. 4.0a (Swofford 2003), using the heuristic search option with 1,000 random sequence addition replicates and tree bisection-reconnection (TBR) with reconnection limit (=8) as the branch swapping algorithm. Max-trees was set at 5,000. Branches collapsed (creating polytomies) if maximum branch length is zero. The Tree Length (TL), Consistency Indices (CI), Retention Indices (RI), Rescaled Consistency Indices (RC) and Homoplasy Index (HI) were calculated for each tree generated. Bayesian Inference (BI) analyses were performed in MrBayes v. 3.2.7a (Ronquist et al. 2012). Six Markov chain Monte Carlo runs were started, and the random start trees were calculated for 50,000,000 generations and sampled every 1,000 generations. 25% of the trees initially produced were discarded as burn-in. ML bootstrap support (MLBS) and MP bootstrap support (PBS) equal or greater than 70% (Hillis and Bull 1993) and Bayesian posterior probabilities (PP) equal or greater than 0.95 (Hespanhol et al. 2019) are displayed on the edited phylogenetic tree. The phylogenetic tree was drawn with FigTree v. 1.4.4 (Rambaut 2009).

Genealogical Phylogenetic Species Recognition (GCPSR) analysis

Morphological and phylogenetically related species were analyzed using the genealogical consistency phylogenetic species identification (GCPSR) model as described by Taylor et al. (2000) by pin-pair homogeneity index test (PHI) (Bruen et al. 2006). The PHI tests were performed in SplitsTree v. 4.17.1 (Huson 1998; Huson and Bryant 2006) as described by Quaedvlieg et al. (2014) to determine the level of recombination within phylogenetically closely related species. The results can be visualized by constructing a split graph using LogDet conversion and the Splits options. The hypothesis of this analysis is if the PHI value is below 0.05 ($\Phi w < 0.05$), there is significant evidence for the presence of recombination.

Results

Phylogenetic analysis

DNA sequences used in this study (Table 1) were selected to obtain phylogenetic trees based on the closest matches by the BLASTn search with strain GUCC 401.6 and eight strains (GUCC 401.1–401.5 to GUCC 401.7–401.9), respectively, with outgroup *C. neolangeronii* (CBS 797.97) and *C. langeronii* (CBS 189.54). The final alignment (GUCC 401.6) of ITS, *tef1* and *act* comprised 1,033 characters, viz. ITS: 1–543, *act*: 544–770 and *tef1*: 771–1033, which included 843 constant characters, 38 variable characters and 152 parsimony-informative characters, and the alignment (GUCC 401.1–GUCC 401.9 except for GUCC 401.6) comprised 1,040 characters, viz., ITS: 1–544, *act*: 545–780 and *tef1*: 781–1040; which included 813 constant characters, 46 variable characters and 181 parsimony-informative characters. The RAxML results were selected to show the topology (Fig. 1 for GUCC 401.6 and Fig. 2 for GUCC 401.1–GUCC 401.9 except for GUCC 401.6), because the MP and Bayesian analyses resulted in similar topologies. The parameter settings that were used are shown in Table 3.

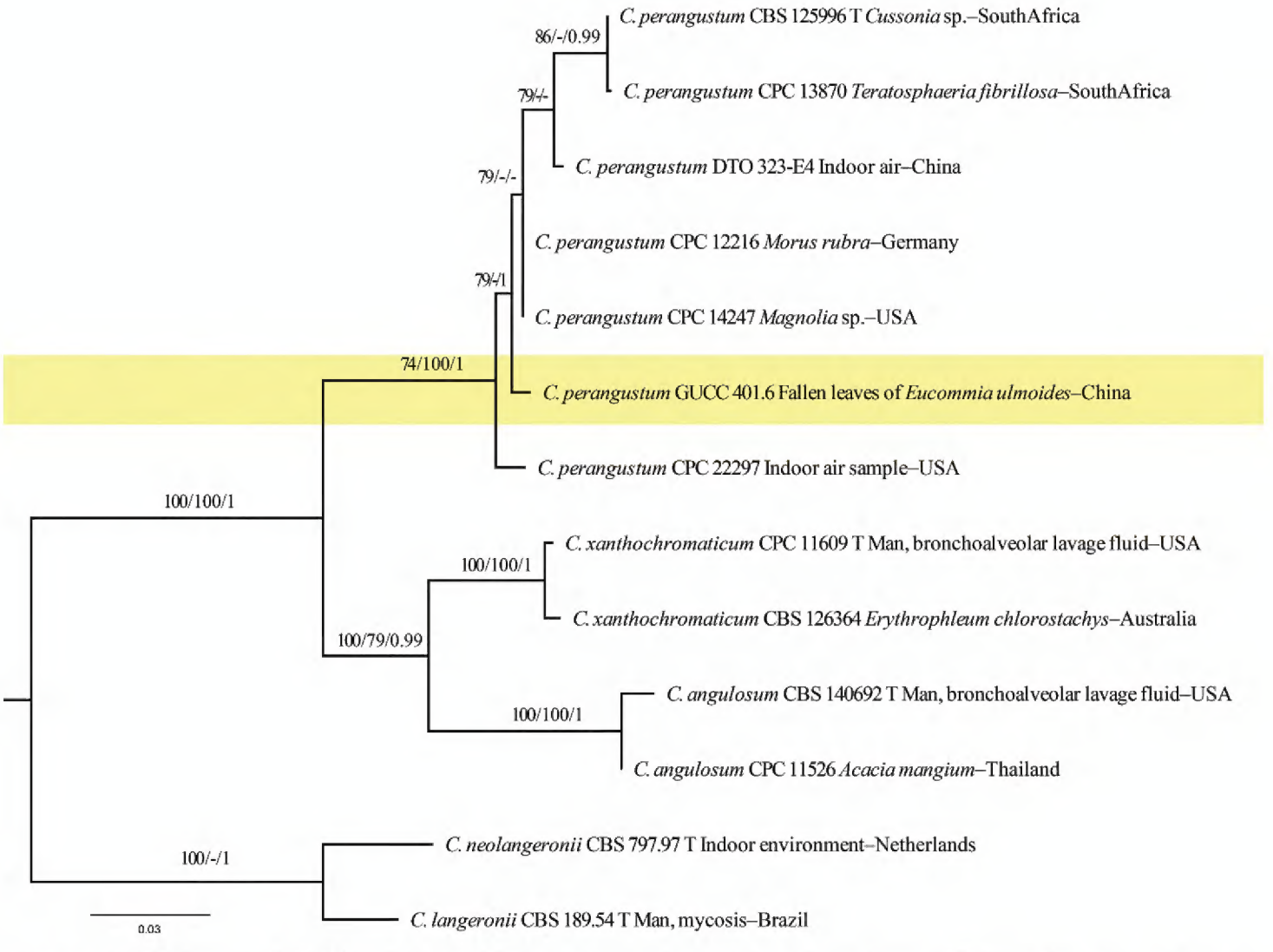
C. perangustum CBS 125996 T Cussonia sp.–SouthAfrica
86/-/0.99
C. perangustum CPC 13870 Teratosphaeria fibrillosa–SouthAfrica
79/-/-
C. perangustum DTO 323-E4 Indoor air–China
79/-/-
C. perangustum CPC 12216 Morus rubra–Germany
79/-/1
C. perangustum CPC 14247 Magnolia sp.–USA
74/100/1
C. perangustum GUCC 401.6 Fallen leaves of Eucommia ulmoides–China
C. perangustum CPC 22297 Indoor air sample–USA
100/100/1
C. xanthochromaticum CPC 11609 T Man, bronchoalveolar lavage fluid–USA
100/100/1
C. xanthochromaticum CBS 126364 Erythrophleum chlorostachys–Australia
100/79/0.99
C. angulosum CBS 140692 T Man, bronchoalveolar lavage fluid–USA
100/100/1
C. angulosum CPC 11526 Acacia mangium–Thailand
C. neolangeronii CBS 797.97 T Indoor environment–Netherlands
100/-/1
C. langeronii CBS 189.54 T Man, mycosis–Brazil
0.03

Figure 1. Maximum Likelihood (RAxML) tree from the combined analysis of ITS, *tef1* and *act* sequences of *Cladosporium*, which includes our strain GUCC 401.6. The tree was rooted with *C. neolangeronii* (CBS 797.97) and *C. langeronii* (CBS 189.54). ML and MP bootstrap values (≥ 70%) and Bayesian posterior probability (≥ 0.95) are indicated along branches (ML/MP/PP). Our species is highlighted with a yellow background. T = ex-holotype strain.

GUCC 401.6 clustered very close to *C. perangustum* (CBS 125996 = ex-holotype strain) with relatively high statistical support (79% MLBS/1 PP) (Fig. 1). Strains GUCC 401.2, GUCC 401.3, GUCC 401.4 and GUCC 401.5 had a very close relationship to *C. tenuissimum* (CBS 125995), variedly supported by MLBS (93%), PBS (70%) and PP (1) (Fig. 2). The comparison of DNA bases (Table 2) showed that our strains cluster with the ex-type strain of *C. tenuissimum* (CBS 125995, ex-epitype strain) with only one base pair difference in the ITS, two to fifteen base pair difference in the *tef1*, and one to five base pair difference in the *act*. *Cladosporium eucommiae* (GUCC 401.1) is a sister

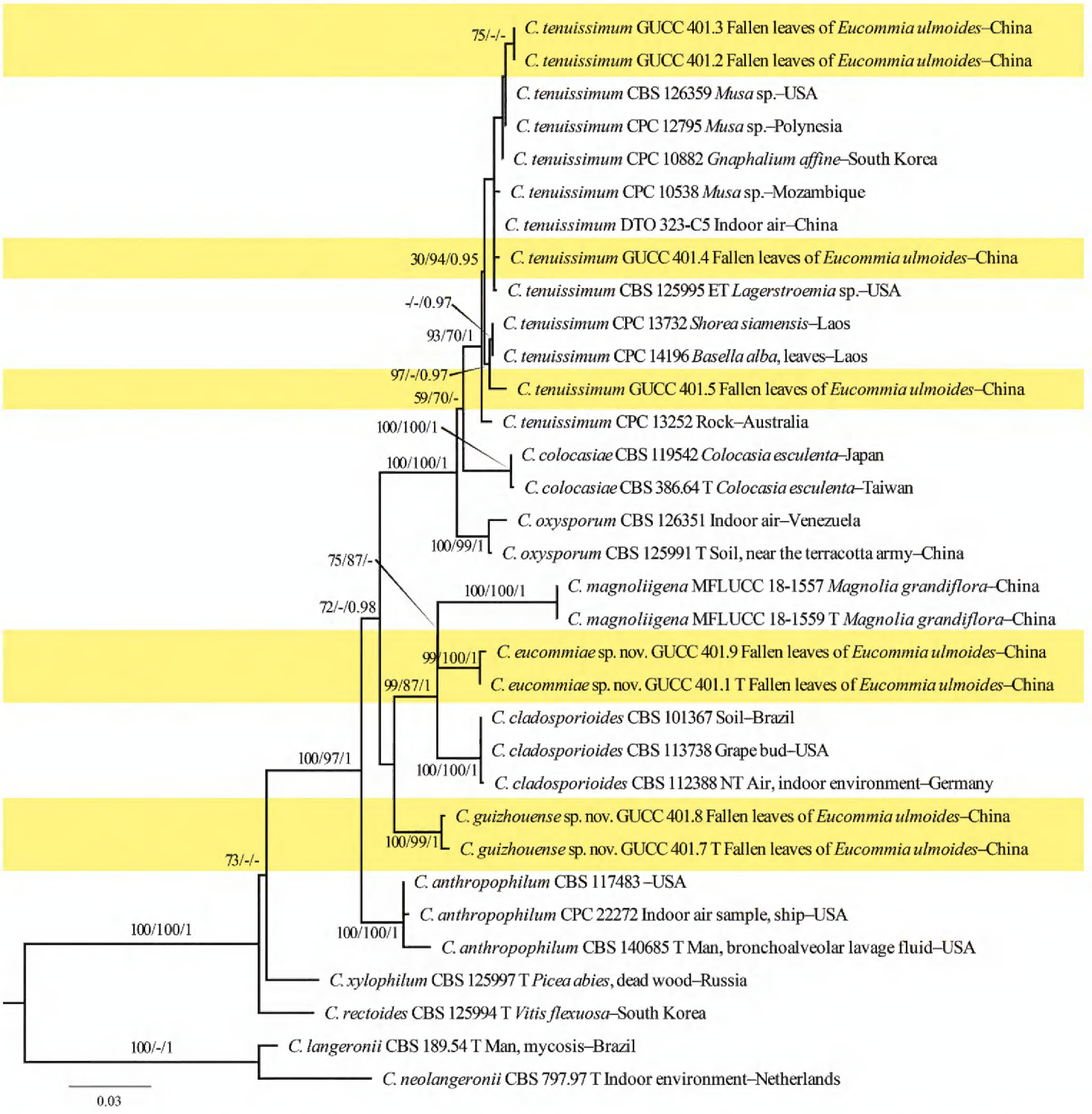
75/-/-
C. tenuissimum GUCC 401.3 Fallen leaves of Eucommia ulmoides–China
C. tenuissimum GUCC 401.2 Fallen leaves of Eucommia ulmoides–China
C. tenuissimum CBS 126359 Musa sp.–USA
C. tenuissimum CPC 12795 Musa sp.–Polynesia
C. tenuissimum CPC 10882 Gnaphalium affine–South Korea
C. tenuissimum CPC 10538 Musa sp.–Mozambique
C. tenuissimum DTO 323-C5 Indoor air–China
30/94/0.95
C. tenuissimum GUCC 401.4 Fallen leaves of Eucommia ulmoides–China
C. tenuissimum CBS 125995 ET Lagerstroemia sp.–USA
-/-/0.97
93/70/1
C. tenuissimum CPC 13732 Shorea siamensis–Laos
C. tenuissimum CPC 14196 Basella alba, leaves–Laos
97/-/0.97
59/70/-
C. tenuissimum GUCC 401.5 Fallen leaves of Eucommia ulmoides–China
100/100/1
C. tenuissimum CPC 13252 Rock–Australia
100/100/1
C. colocasiae CBS 119542 Colocasia esculenta–Japan
C. colocasiae CBS 386.64 T Colocasia esculenta–Taiwan
C. oxysporum CBS 126351 Indoor air–Venezuela
100/99/1
C. oxysporum CBS 125991 T Soil, near the terracotta army–China
75/87/-
100/100/1
C. magnoliigena MFLUCC 18-1557 Magnolia grandiflora–China
72/-/0.98
C. magnoliigena MFLUCC 18-1559 T Magnolia grandiflora–China
99/100/1
C. eucommiae sp. nov. GUCC 401.9 Fallen leaves of Eucommia ulmoides–China
99/87/1
C. eucommiae sp. nov. GUCC 401.1 T Fallen leaves of Eucommia ulmoides–China
C. cladosporioides CBS 101367 Soil–Brazil
C. cladosporioides CBS 113738 Grape bud–USA
100/97/1
100/100/1
C. cladosporioides CBS 112388 NT Air, indoor environment–Germany
C. guizhouense sp. nov. GUCC 401.8 Fallen leaves of Eucommia ulmoides–China
100/99/1
C. guizhouense sp. nov. GUCC 401.7 T Fallen leaves of Eucommia ulmoides–China
73/-/-
C. anthropophilum CBS 117483 –USA
C. anthropophilum CPC 22272 Indoor air sample, ship–USA
100/100/1
100/100/1
C. anthropophilum CBS 140685 T Man, bronchoalveolar lavage fluid–USA
C. xylophilum CBS 125997 T Picea abies, dead wood–Russia
C. rectoides CBS 125994 T Vitis flexuosa–South Korea
100/-/1
C. langeronii CBS 189.54 T Man, mycosis–Brazil
C. neolangeronii CBS 797.97 T Indoor environment–Netherlands
0.03

Figure 2. Maximum Likelihood (RAxML) tree from the combined analysis of ITS, *tef1* and *act* sequences of *Cladosporium*, which includes our strains GUCC 401.1–GUCC 401.9 (except for GUCC 401.6). The tree was rooted with *C. neolangeronii* (CBS 797.97) and *C. langeronii* (CBS 189.54). ML and MP bootstrap values (≥ 70%) and Bayesian posterior probability (≥ 0.95) are indicated along branches (ML/MP/PP). Our species are emphasized with a yellow background. T = ex-holotype strain, ET = ex-epitype strain, NT = ex-neotype strain.

to *C. magnoliigena* (MFLUCC 18-1559) and *C. cladosporioides* (CBS 101367) with high statistical support (75% MLBS / 87% MPBS)/(99 MLBS / 87% MPBS / 1 PP) (Fig. 2). The comparison of DNA bases composition (Table 2) indicated that, between *C. eucommiae* (GUCC 401.1) and *C. magnoliigena*, there were identical sequences in the ITS region, but 29 bases different in the *tef1* region. Unfortunately, *Cladosporium magnoliigena* did not have *act* sequence data for comparison. The comparison of DNA bases composition (Table 2) indicated that, between *C. eucommiae* (GUCC 401.1) and *C. cladosporioides* (CBS 112388, ex-neotype strain), there were 18 bp differences in the *tef1* region, and 13 in the *act* region, but without difference in the ITS sequences. GUCC 401.7 was closer to *C. cladosporioides* (CBS 112388, ex-neotype strain) with high support in their respective branches (100% MLBS / 99% MPBS / 1 PP)/(100% MLBS / 100% MPBS / 1 PP) (Fig. 2). The comparison of DNA bases (Table 2) reveals 29 bp difference on *tef1* and 14 bp difference in *act* between *C. guizhouense* and *C. cladosporioides* (CBS 112388, ex-neotype strain), but only 1 bp difference in ITS sequences.

The pairwise homoplasy index (PHI) test revealed that there was no significant recombination (Φw = 0.4589) between *C. eucommiae* (GUCC 401.1 and GUCC 401.9) and the related taxa *C. magnoliigena*, *C. cladosporioides*, *C. guizhouense* (GUCC

Table 2. The DNA base differences between our strains and related taxa in the three gene regions. Asterisks (*) denote our material.

Species	Strain number	Gene region and alignment positions		
		ITS (1–489 characters)	***tef1* (490–718 characters)**	***act* (719–1008 characters)**
C. eucommiae sp. nov.*	GUCC 401.1^{T}	–	–	–
C. eucommiae sp. nov.*	GUCC 401.9	0	3	0
C. magnoliigena	MFLUCC 18-1559^{T}	0	29	n/a
C. magnoliigena	MFLUCC 18-1557	0	29	n/a
C. cladosporioides	CBS 112388	0	18	13
C. cladosporioides	CBS 113738	0	16	13
C. cladosporioides	CBS 101367	1	16	13
		ITS (1–542 characters)	***tef1* (543–796 characters)**	***act* (797–1029 characters)**
C. tenuissimum	CBS 125995^{ET}	–	–	–
*C. tenuissimum**	GUCC 401.2	0	3	5
*C. tenuissimum**	GUCC 401.3	0	15	n/a
*C. tenuissimum**	GUCC 401.4	0	2	1
*C. tenuissimum**	GUCC 401.5	1	9	3
		ITS (1–507 characters)	***tef1* (508–743 characters)**	***act* (744–948 characters)**
*C. perangustum**	GUCC 401.6	–	–	–
C. perangustum	CBS 12599^{6}T	0	26	7
C. perangustum	CPC 13870	0	22	7
C. perangustum	DTO 323-E4	0	13	5
C. perangustum	CPC 12216	0	2	5
C. perangustum	CPC 14247	0	2	5
		ITS (1–480 characters)	***tef1* (481–728 characters)**	***act* (729–933 characters)**
C. guizhouense sp. nov.*	GUCC 401.7^{T}	–	–	–
C. guizhouense sp. nov.*	GUCC 401.8	0	3	2
C. cladosporioides	CBS 112388	1	29	14
C. cladosporioides	CBS 113738	1	27	14
C. cladosporioides	CBS 101367	2	27	14

Table 3. The parameters of MP and Bayesian methods in this study.

Strain number	MP						Bayesian		
	TL	PT	CI	RI	RC	HI	Model	Unique site patterns	ASDSF
GUCC 401.1 –GUCC 401.9 (except for GUCC 401.6)	400	300	0.7475	0.8648	0.6464	0.2525	ITS: JC+I; *tef1*: GTR+G; *act*: HKY+G	Division 1 = 54 Division 2 = 99 Division 3 = 154	0.009875
GUCC 401.6	281	2	0.8505	0.8817	0.7499	0.1495	ITS: SYM; *tef1*: GTR+G; *act*: GTR+G	Division 1 = 30 Division 2 = 75 Division 3 = 128	0.009961

TL: Tree length; PT: Parsimonious tree; CI: Consistency Indices; RI: Retention Indices; RC: Rescaled Consistency Indices HI: Homoplasy Index; Model: the models used for the different partitions; ASDSF: average standard deviation of split frequencies.

401.7 and GUCC 401.8). The PHI test did not find any statistically significant evidence for recombination (Φw = 0.02487) between our four strains (GUCC 401.2, GUCC 401.3, GUCC 401.4 and GUCC 401.5) and the related taxon *C. tenuissimum* (CBS 126359, ex-epitype strain, CPC 12795, CPC 10882, CPC 10538, DTO 323-C5, CBS 125995, CPC 13732, CPC 14196 and CPC 13252). Based on the PHI test, there was a statistically significant recombination (Φw = 0.0104) between GUCC 401.6 and the related taxon *C. perangustum* (CBS 125996, = ex-holotype strain, CPC 13870, DTO 323-E4, CPC 12216, CPC 14247 and CPC 22297).

Taxonomy

In this section, we introduced two new species and report two new substrate records.

Cladosporium eucommiae **S.Y. Wang, Yong Wang bis & Y. Li, sp. nov.**
MycoBank No: 842406
Fig. 3a–h

Etymology. *eucommiae*, in reference to the genus name of the host plant (*Eucommia ulmoides*), from which the fungus was isolated.

Type. China, Guizhou Province, Guiyang, Huaxi district, plantation forest of *E. ulmoides*, Guizhou University (26°24'16"N, 106°40'29"E), on fallen leaves of *E. ulmoides*, S.Y. Wang, Y. Wang & Y. Li, 13 January 2021 (HGUP 401.1, ***holotype***; ex-type living culture GUCC 401.1; additional living culture GUCC 401.9).

Description. Saprobic on fallen leaves of *Eucommia ulmoides*. **Sexual morph:** Not developed. **Asexual morph**: Hyphomycetous. ***Mycelium*** abundant, superficially and submerged, overgrowing whole culture dishes, thin to dense, hyphae straight to slightly sinuous, branched, light olive-green to olive-brown, 1.5–5 µm wide, thin-walled, smooth. ***Conidiophores*** (7–)22–198 × 2.5–4.5 µm ($\bar{x}$ = 77.2 × 3.3 µm; n = 20), erect, branching, slightly attenuated towards the apex, light olive-green, smooth and thick-walled. ***Conidia*** 3–9 × 2.5–4.5 µm ($\bar{x}$ = 5.6 × 3.3 µm; n = 30), in simple and branched acropetal chains, mostly light olive, aseptate, smooth-walled and thin-walled, variable in size and shape, subglobose, ellipsoid-ovoid, obovoid, fusiform, subcylindrical.

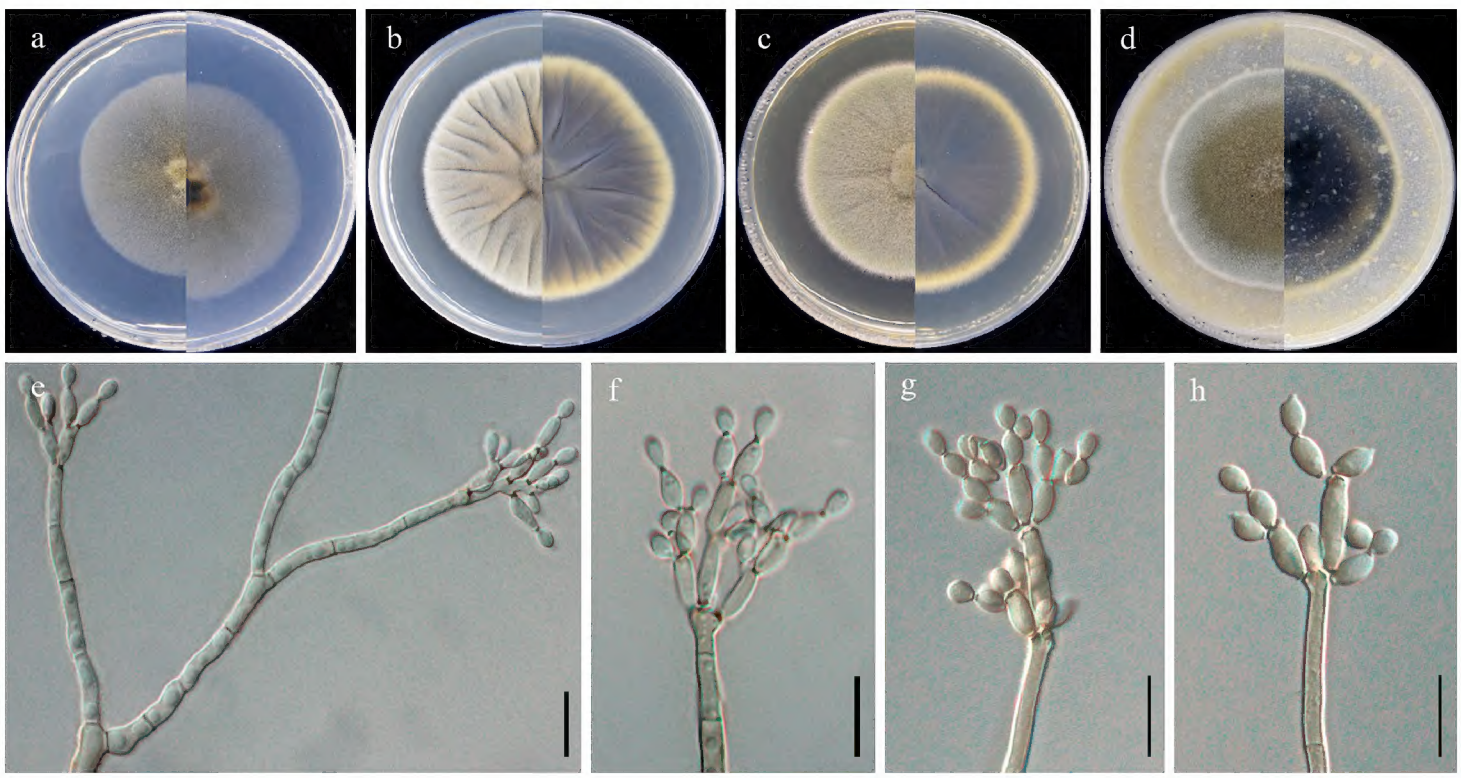

Figure 3. *Cladosporium eucommiae* (GUCC 401.1, ex holotype strain). **a–d** colonies on SNA, PDA, MEA and OA (left: above, right: reverse) **e** branching conidiophore, secondary ramoconidia and conidia on SNA **f–h** conidiogenous cells, secondary ramoconidia and conidia on SNA. Scale bars: 10 μm (**e–h**).

Secondary ramoconidia 5–25 × 2.5–4.0 μm ($\overline{x}$ = 11.9 × 3.4 μm; n = 30), olive-green, ellipsoid-ovoid, obovoid, fusiform, subcylindrical, aseptate, smooth-walled and thin-walled, rarely thick-walled.

Culture characteristics. ***Colonies*** on SNA 35–45 mm diam, after 2 weeks at 25 °C, pale olive, flat, velvety, with a regular edge, reverse light olive. ***Colonies*** on PDA 30–45 mm diam, after 2 weeks at 25 °C, olive-brown to gray-olive to iron-gray, with a regular white edge, irregularly folded, slightly depressed at the center, thatched, and often forming a bulge in the colony kernel, reverse olive to dark olive, with a whitish final edge. ***Colonies*** on MEA 35–45 mm diam, after 2 weeks at 25 °C, gray-green to olive, less radially furrowed, velvety, with an even gray white edge, reverse olive to dark olive, with an even gray-green final edge. ***Colonies*** on OA 35–40 mm diam, after 2 weeks at 25 °C, olive to gray-green, white at the final edge, flat, velvety, margin regular; reverse dark green to black, with a whitish final edge.

GenBank numbers. ITS: OL587465, *tef1*: OL504966, *act*: OL519775 (GUCC 401.1); ITS: ON334729, *act*: ON383337 (GUCC 401.9).

Cladosporium guizhouense **S.Y. Wang, Yong Wang bis & Y. Li, sp. nov.**
MycoBank No: 842407
Fig. 4a–h

Etymology. guizhouense, in reference to the type location (Guizhou Province), where the fungus was isolated.

Type. China, Guizhou Province, Guiyang, Huaxi district, plantation forest of *Eucommia ulmoides*, Guizhou University (26°24'16"N, 106°40'29"E), on fallen leaves of *E. ulmoides*, S.Y. Wang, Y. Wang & Y. Li, 13 January 2021 (HGUP 401.6, ***holotype***; living culture GUCC 401.7; additional living culture GUCC 401.8).

Description. Saprobic on fallen leaves of *Eucommia ulmoides*. **Sexual morph**: Not developed. **Asexual morph**: Hyphomycetous. ***Mycelium*** abundant, submerged, overgrowing whole culture dishes, hyphae straight to slightly sinuous, septate, branching, light olive-green to olive-brown, mostly smooth- and thin-walled, 1.5–6 µm wide. ***Conidiophores*** 13–100 × 3–4.5 µm ($\bar{x}$ = 60.8 × 3.5 µm; n = 10), erect, branching, light olive-green, smooth- and thin-walled. ***Conidia*** 3–7.5 × 2.5–4 µm ($\bar{x}$ = 4.8 × 3.1 µm; n = 30), in simple and branched acropetal chains, mostly light olive, aseptate, mostly smooth- and thin-walled, variable in size and shape, ellipsoid-ovoid, obovoid, fusiform. ***Secondary ramoconidia*** 6.5–23 × 3–5.5 µm ($\bar{x}$ = 11.3 × 4.1 µm; n = 30), pale olive-green, narrowly ellipsoid to cylindrical-oblong, subcylindrical, aseptate, smooth- and thin-walled.

Culture characteristics: ***Colonies*** on SNA 45–55 mm diam, after 2 weeks at 25 °C, pale olive, flat, velvety, margin regularly, reverse light olive. ***Colonies*** on PDA 40–50 mm diam, after 2 weeks at 25 °C, smoke-gray to light olive-gray, reverse leaden-gray, gray-olive at edge both surface and reverse, woolly or felty, broad edge, regular, growth low convex, without protuberant exudates, reverse formed cracks in the middle small circle. ***Colonies*** on MEA 30–40 mm diam, after 2 weeks at 25 °C, smoke-gray to light olive-gray, woolly or felty, fluffy, with a whitish narrow final edge; reverse olive-yellow or olive-brown, radially furrowed, irregularly folded, with

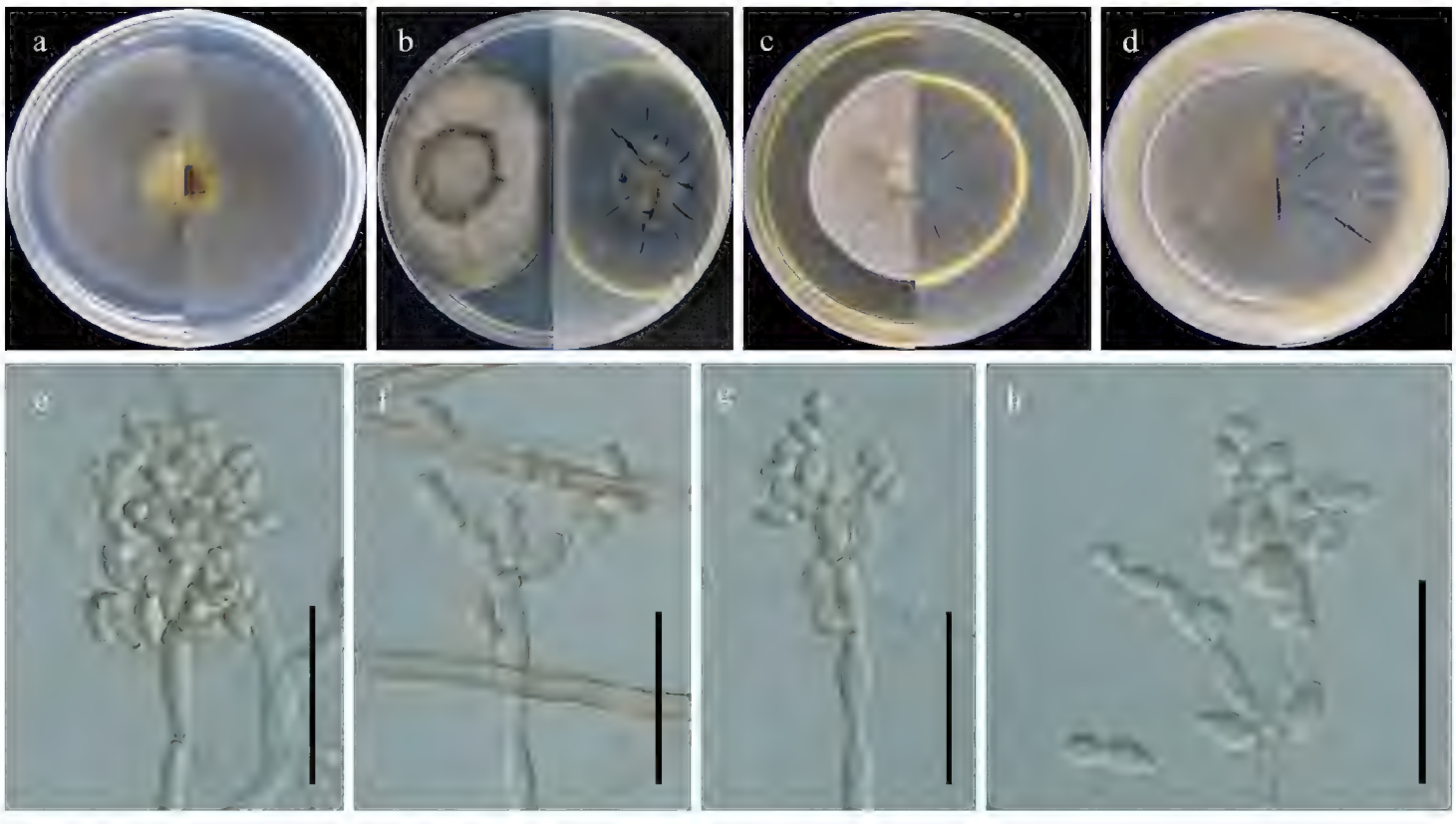

Figure 4. *Cladosporium guizhouense* (GUCC 401.7). **a–d** colony on SNA, PDA, MEA and OA (left: above, right: reverse) **e–h** conidiogenous cells, secondary ramoconidia and conidia on SNA. Scale bars: 10 µm (**e–h**).

a whitish narrow final edge. ***Colonies*** on OA 30–45 mm diam, after 2 weeks at 25 °C, gray-green or olive, granular and fluffy mycelium, woolly and felty edge, with an irregularly folded whitish and olive final edge; reverse olive-yellow or olive-brown, with a whitish narrow final edge.

GenBank numbers. ITS: OL579741, *tef1*: OL504965, *act*: OL519780 (GUCC 401.7); ITS: ON334728, *tef1*: ON383470, *act*: ON383338 (GUCC 401.8).

Cladosporium perangustum **Bensch, Crous & U. Braun, Studies in Mycology 67: 65 (2010)**

MycoBank No: 517085

Fig. 5a–h

Material examined. China, Guizhou Province, Guiyang, Huaxi district, plantation forest of *E. ulmoides*, Guizhou University (26°24'16"N, 106°40'29"E), on fallen leaves of *E. ulmoides*, S.Y. Wang, Y. Wang & Y. Li, 13 January 2021 (HGUP 401.6, living culture GUCC 401.6) (new substrate record).

Description. Saprobic on fallen leaves of *Eucommia ulmoides.* **Sexual morph**: Not developed. **Asexual morph**: Hyphomycetous. ***Mycelium*** superficial, hyphae branched, hyaline to subhyaline, 2.5–5 µm wide, usually slightly constricted at the septa and some-

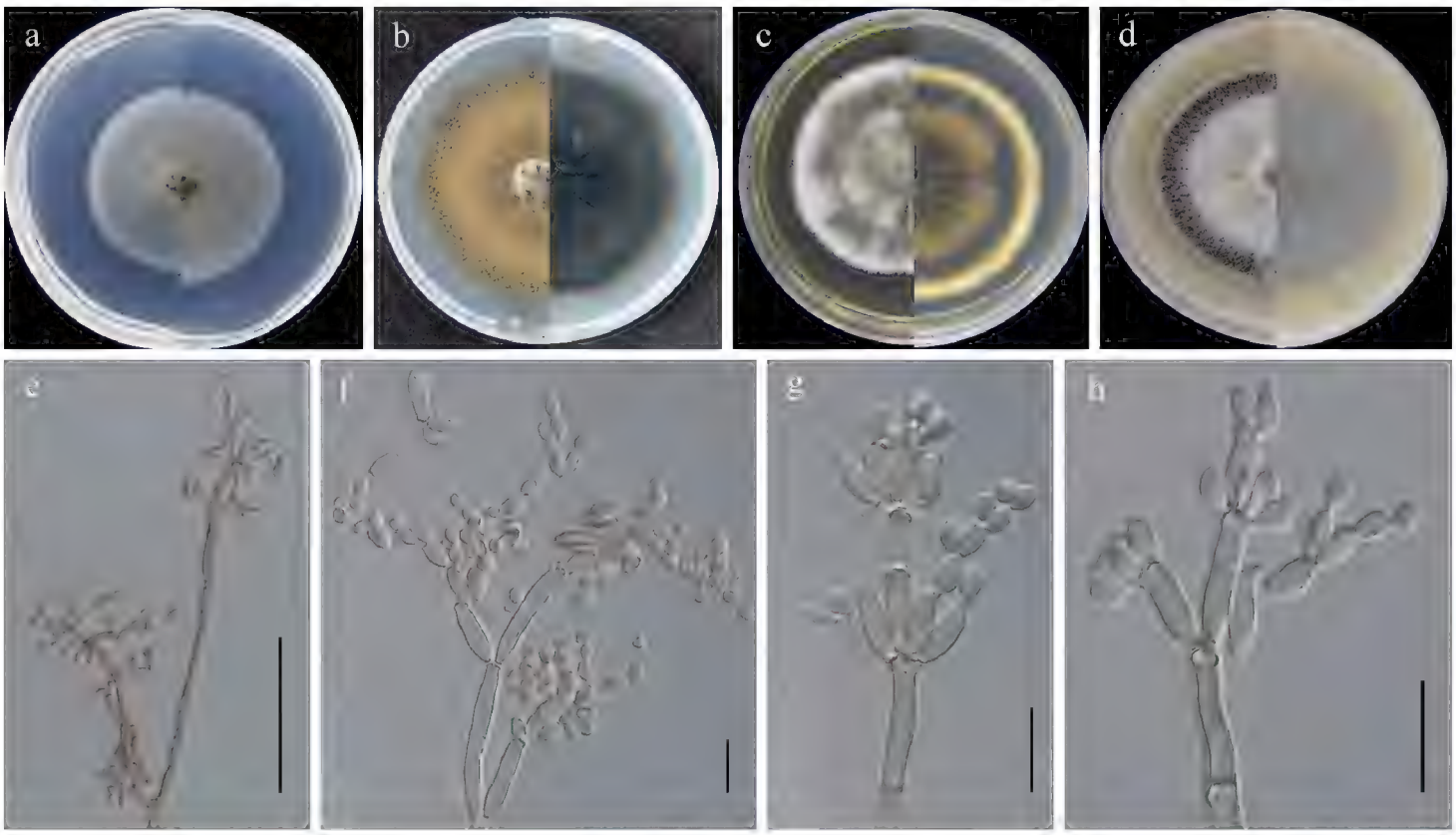

Figure 5. *Cladosporium perangustum* (GUCC 401.6, new substrate record from Guizhou Province). **a–d** colonies on SNA, PDA, MEA and OA (left: above, right: reverse) **e** branching conidiophore, secondary ramoconidia and conidia on SNA **f–h** conidiogenous cells, secondary ramoconidia and conidia on SNA. Scale bars: 50 µm (**e**); 10 µm (**f–h**).

what swollen, smooth to somewhat verruculose or irregularly rough-walled. ***Conidiophores*** macro- and micronematous, 14–167 × 2.5–4.5 µm ($\bar{x}$ = 65.4 × 3.4 µm; n = 20), erect, branched, slightly attenuated towards the apex, light olive-green, smooth and thick-walled. ***Conidia*** in acropetal chains, 2–9.5 × 2–4 µm ($\bar{x}$ = 5.6 × 3.3 µm; n = 30) mostly light olive-green, aseptate, mostly smooth-walled and thin-walled, variable in size and shape, subglobose, ellipsoid-ovoid, obovoid, fusiform. ***Secondary ramoconidia*** 6–24 × 2–5.5 µm ($\bar{x}$ = 11.2 × 3.3 µm; n = 30), olive-green, narrowly ellipsoid to cylindrical-oblong, subcylindrical, aseptate, rarely 1-septate, mostly smooth-walled and thick-walled.

Culture characteristics. ***Colonies*** on SNA 30–40 mm diam, after 2 weeks at 25 °C, pale olive to pale whitish, flat, velvety, with a regular edge, reverse light olive to light white. ***Colonies*** on PDA 30–40 mm diam, after 2 weeks at 25 °C, olive-gray to olive-green or olive-brown, powdery or flocculent, fluffy, regular, radially furrowed, lacerated or feathery, and often forming a gray-white or olive bulge in the colony kernel; reverse dark olive or dull green to black. ***Colonies*** on MEA 35–45 mm diam, after 2 weeks at 25 °C, gray-green to white or gray-white, fluffy, radially furrowed, with a whitish final edge; reverse olive-yellow to olive-gray to olive-green, with a whitish final edge. ***Colonies*** on OA 35–45 mm diam, after 2 weeks at 25°C, white to olive-green, with a pale gray final edge, velvety or fluffy, margins colorless or pale gray, glabrous, regular; reverse olive-green to dark green.

GenBank numbers. ITS: OL579742, *tef1*: OL581726, *act*: OL519779.

Cladosporium tenuissimum **Cooke, Grevillea 6: 140 (1878)**
MycoBank No: 145672
Fig. 6a–i

Materials examined. China, Guizhou Province, Guiyang, Huaxi district, plantation forest of *E. ulmoides*, Guizhou University (26°24'16"N, 106°40'29"E), on fallen leaves of *E. ulmoides*, S.Y. Wang, Y. Wang & Y. Li, 13 January 2021, (HGUP 401.1; HGUP 401.2; HGUP 401.3 and HGUP 401.4, living cultures GUCC 401.2; GUCC 401.3; GUCC 401.4 and GUCC 401.5) (new substrate record).

Description. Saprobic on fallen leaves of *Eucommia ulmoides.* **Sexual morph**: Not developed. **Asexual morph**: Hyphomycetous. ***Mycelium*** abundant, superficial and submerged, overgrowing whole culture dishes, hyphae straight to slightly sinuous, septate, branching, light olive-green to olive-brown, smooth-walled, 1.5–6 µm wide. ***Conidiophores*** 13–100 × 2.5–4.5 µm ($\bar{x}$ = 60.8 × 3.6 µm; n = 10), erect, branching, light olive-green, smooth and thin walled. ***Conidia*** 2.5–7.5 × 2–4 µm ($\bar{x}$ = 4.9 × 3.2 µm; n = 30), in simple and branched acropetal chains, mostly light olive-green, aseptate, mostly smooth- and thin-walled, variable in size and shape, subglobose, ellipsoid-ovoid, obovoid, fusiform. ***Secondary ramoconidia*** 5.5–23 × 2.5–5.5 µm ($\bar{x}$ = 0.9 × 3.8 µm; n = 30), pale olive-green, narrowly ellipsoid to cylindrical-oblong or subcylindrical, sometimes septate and sometimes aspetate (1-septate appear at maturity), smooth- and thin-walled.

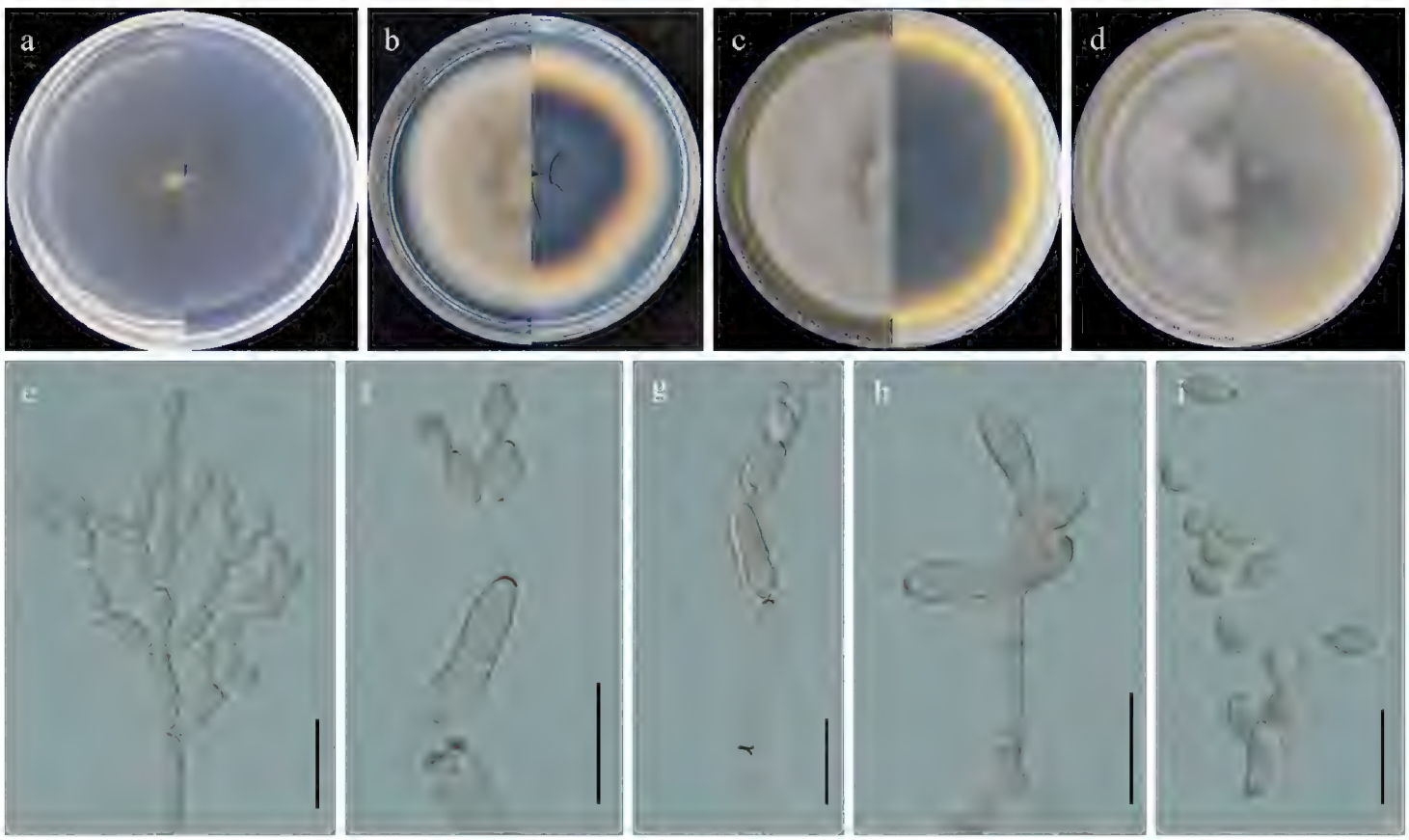

Figure 6. *Cladosporium tenuissimum* (GUCC 401.2, new substrate record from Guizhou Province). **a–d** colonies on SNA, PDA, MEA and OA (left: above, right: reverse) **e–h** secondary ramoconidia and conidia on SNA **i** conidia on SNA. Scale bars: 10 μm (**e–i**).

Culture characteristics: ***Colonies*** on SNA 50–55 mm diam, after 2 weeks at 25 °C, pale olive to pale white, flat, velvety, with a regular edge, reverse light olive to light white. ***Colonies*** on PDA 40–55 mm diam, after 2 weeks at 25 °C, smoke-gray to light olive-gray or olive to light olive-gray, reverse leaden-gray, gray-olive at edge both surface and reverse, woolly or felty, broad edge, regular, growth low convex, without protuberant exudates, occasionally reverse formed a sunflower like shape in the middle. ***Colonies*** on MEA 40–50 mm diam, after 2 weeks at 25 °C, olive-gray or gray, fluffy; reverse olive-green to dark olive, with an olive-yellow to gray-white edge, radially furrowed. ***Colonies*** on OA 40–60 mm diam, after 2 weeks at 25 °C, gray-white or iron-gray to gray-olive, fluffy to felty; reverse olive-brown to olive.

GenBank numbers. ITS: OL579746, *tef1*: OL504967, *act*: OL519776 (GUCC 401.2); ITS: OL579745, *tef1*: OL505077 (GUCC 401.3); ITS: OL579744, *tef1*: OL581724, *act*: OL519777 (GUCC 401.4); ITS: OL579743, *tef1*: OL581725, *act*: OL519778 (GUCC 401.5).

Discussion

In this paper, we revealed four *Cladosporium* taxa on fallen leaves of *E. ulmoides*, two of which are described here as new to science. Phylogenetic analyses showed that *C. eucommiae* is different from *C. magnoliigena* (Jayasiri et al. 2019), although

act sequences are not available for the latter species. Conidia of *C. eucommiae* (3–9 × 2.5–4.5 μm) are usually narrower and longer than those of *C. magnoliigena* (4.2–5.5 × 2–5 μm), while secondary ramoconidia of *C. eucommiae* are usually aseptate and longer than those of *C. magnoliigena* (5–25 × 2.5–4.0 μm *vs* 9.5–18 × 2.7–4.2 μm and 0–3-septate). Thus, the two species are clearly distinct in morphology as well as DNA sequence data. Phylogenetic analyses showed that sequences retrieved from GUCC 401.7 and GUCC 401.8 are different from those obtained from *C. cladosporioides* (CBS 112388, ex-neotype strain) (Bensch et al. 2010) by phylogenetic analyses (Fig. 2). Conidia of GUCC 401.7 and *C. cladosporioides* show no significant differences in size, color and shape, but secondary ramoconidia of GUCC 401.7 were usually shorter than those of *C. cladosporioides* (6.5–23 × 3–5.5 μm *vs* 15–50 × (2.5–)3–5 μm), and conidiophores of GUCC 401.7 (13–100 × 3–4.5 μm) were shorter than in *C. cladosporioides* (40–300(–350) × (2.5–)3–4(–5.5) μm). Therefore, there are significant differences in the morphology and DNA sequence data between the two species. The combination of morphology, phylogenetic analyses, comparison of DNA base composition and GCPSR analysis support our proposal that *C. eucommiae* and *C. guizhouense* represent two novel taxa.

Sequences retrieved from GUCC 401.6 clustered among six sequences obtained from *C. perangustum* strains (Fig. 1), but conidia of GUCC 401.6 (2–9.5 × 2–4 μm) were usually somewhat narrower and longer than CBS 125996 (Bensch et al. 2010) (2–4(–5) × (1.5–)2–2.5 μm), and secondary ramoconidia of GUCC 401.6 (6–24 × 2–5.5 μm) were wider than those of *C. perangustum* (6–30(–34) × 2–3(–3.5) μm). In addition, GUCC 401.6 can be well distinguished from *C. perangustum* by its slower growing colonies in PDA, MEA and OA (30–40, 35–45 and 35–45 mm diam/14 d), whereas CBS 125996 grew 33–76, 40–72 and 40–75 mm diam/14 d. Although morphology and phylogeny showed minor differences, GCPSR analysis supported statistically significant recombination, after careful consideration, GUCC 401.6 was identified as *C. perangustum.* The differences may be caused by different substrates or geographical regions, which needs further investigation. Conidiophores of GUCC 401.2–GUCC 401.5 were shorter than in CBS 125995 (13–100 × 2.5–4.5 μm *vs* 49–542(–800) × (3–)4–7 μm), but secondary ramoconidia and conidia (5.5–23 × 2.5–5.5 μm; 2.5–7.5 × 2–4 μm) were similar to those of *C. tenuissimum* (15–31 × 4–5 μm; 3–13 × 2–6 μm) (Cooke 1878). Sequences retrieved from our four strains cluster with sequences obtained from *C. tenuissimum* strains (Fig. 2) with minor DNA base differences. Thus, our four strains were identified as *C. tenuissimum.*

Our five strains pertain to two known species, viz., *C. perangustum* and *C. tenuissimum*, but with *E. ulmoides* as new substrate records for these species. The main focus of this study was the exploration of the diversity of microfungi associated with a *E. ulmoides* plantation forest. In previous studies, *Cladosporium parapenidielloides* was found on *Eucalyptus* sp. in Australia, *C. perangustum* on *Magnolia* sp. in the USA, and *C. pini-ponderosae* on *Pinus ponderosa* in Argentina. So far, *Cladosporium* species have never been isolated from fallen leaves of *E. ulmoides*, the only species of the genus *Eucommia*.

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References

Abdollahzadeh J, Groenewald JZ, Coetzee M, Wingfield MJ, Crous PW (2020) Evolution of lifestyles in Capnodiales. Studies in Mycology 95: 381–414. https://doi.org/10.1016/j.simyco.2020.02.004

Bensch K, Groenewald JZ, Dijksterhuis J, Starink-Willemse M, Andersen B, Summerell BA, Shin H-D, Dugan FM, Schroers H-J, Braun U, Crous PW (2010) Species and ecological diversity within the *Cladosporium cladosporioides* complex (Davidiellaceae, Capnodiales). Studies in Mycology 67: 1–94. https://doi.org/10.3114/sim.2010.67.01

Bensch K, Braun U, Groenewald JZ, Crous PW (2012) The genus *Cladosporium*. Studies in Mycology 72: 1–401. https://doi.org/10.3114/sim0003

Bensch K, Groenewald JZ, Braun U, Dijksterhuis J, Yanez-Morales M, Crous PW (2015) Common but different: The expanding realm of *Cladosporium*. Studies in Mycology 82(1): 23–74. https://doi.org/10.1016/j.simyco.2015.10.001

Bensch K, Groenewald JZ, Meijer M, Dijksterhuis J, Jurjevic Z, Andersen B, Houbraken J, Crous PW, Samson RA (2018) *Cladosporium* species in indoor environments. Studies in Mycology 89(1): 177–301. https://doi.org/10.1016/j.simyco.2018.03.002

Brown KB, Hyde KD, Guest DI (1998) Preliminary studies on endophytic fungal communities of *Musa acuminata* species complex in Hong Kong and Australia. Fungal Diversity 1: 27–51.

Bruen TC, Philippe H, Bryant D (2006) A simple and robust statistical test for detecting the presence of recombination. Genetics 172(4): 2665–2681. https://doi.org/10.1534/genetics.105.048975

Carbone I, Kohn LM (1999) A method for designing primer sets for speciation studies in filamentous Ascomycetes. Mycologia 91(3): 553–556. https://doi.org/10.1080/00275514.1999.12061051

Chomnunti P, Hongsanan S, Aguirre-Hudson B, Tian Q, Persoh D, Dhami MK, Alias AS, Xu JC, Liu XZ, Stadler M, Hyde KD (2014) The sooty moulds. Fungal Diversity 66(1): 1–36. https://doi.org/10.1007/s13225-014-0278-5

Chung D, Kim H, Choi HS (2019) Fungi in salterns. Journal of Microbiology 57(9): 717–724. https://doi.org/10.1007/s12275-019-9195-3

Cooke MC (1878) Ravenel's American fungi. Grevillea 6: 129–146.

Cronquist A (1981) An integrated system of classification of flowering plants. Columbia University Press, New York. https://doi.org/10.2307/2806386

David JC (1997) A contribution to the systematics of *Cladosporium*. Revision of the fungi previously referred to *Heterosporium*. Mycological Papers 172: 1–157.

El-Morsy EM (2000) Fungi isolated from the endorhizosphere of halophytic plants from the Red Sea Coast of Egypt. Fungal Diversity 5: 43–54.

Ellis M (1977) More Dematiaceous Hyphomycetes. Commonwealth Mycological Institute, Kew, Surrey, England. https://doi.org/10.2307/3758674

Hall TA (1999) BioEdit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series 41: 95–98. https://doi.org/10.1021/bk-1999-0734.ch008

Halo BA, Maharachchikumbura SS, Al-Yahyai RA, Al-Sadi AM (2019) *Cladosporium omanense*, a new endophytic species from *Zygophyllum coccineum* in Oman. Phytotaxa 388(1): 145–154. https://doi.org/10.11646/phytotaxa.388.1.8

Hespanhol L, Vallio CS, Costa LM, Saragiotto BT (2019) Understanding and interpreting confidence and credible intervals around effect estimates. Brazilian Journal of Physical Therapy 23(4): 290–301. https://doi.org/10.1016/j.bjpt.2018.12.006

Hillis DM, Bull JJ (1993) An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. Systematic Biology 42(2): 182–192. https://doi.org/10.1093/sysbio/42.2.182

Huson DH (1998) SplitsTree: Analyzing and visualizing evolutionary data. Bioinformatics 14(1): 68–73. https://doi.org/10.1093/bioinformatics/14.1.68

Huson DH, Bryant D (2006) Application of phylogenetic networks in evolutionary studies. Molecular Biology and Evolution 23(2): 254–267. https://doi.org/10.1093/molbev/msj030

Isabel IG, Dania G, Josepa G (2021) Novel species of *Cladosporium* from environmental sources in Spain. MycoKeys 77: 1–25. https://doi.org/10.3897/mycokeys.77.60862

Jayasiri SC, Hyde KD, Jones EBG, Mckenzie E, Karunarathna SC (2019) Diversity, morphology and molecular phylogeny of Dothideomycetes on decaying wild seed pods and fruits. Mycosphere 10(1): 1–186. https://doi.org/10.5943/mycosphere/10/1/1

Katoh K, Standley DM (2016) A simple method to control over-alignment in the MAFFT multiple sequence alignment program. Bioinformatics 32(13): 1933–1942. https://doi.org/10.1093/bioinformatics/btw108

Larsson A (2014) AliView: A fast and lightweight alignment viewer and editor for large datasets. Bioinformatics 22(22): 3276–3278. https://doi.org/10.1093/bioinformatics/btu531

Liang Y, Ran SF, Bhat J, Hyde KD, Wang Y, Zhao DG (2018) *Curvularia microspora* sp. nov. associated with leaf diseases of *Hippeastrum striatum* in China. MycoKeys 29: 49–61. https://doi.org/10.3897/mycokeys.29.21122

Link HF (1816) Observationes in ordines plantarum naturales. 2. Magazin der Gesellschaft Naturforschenden Freunde Berlin 8: 25–45.

Marin-Felix Y, Groenewald JZ, Cai L, Chen Q, Marincowitz S, Barnes I, Bensch K, Braun U, Camporesi E, Damm U, de Beer ZW, Dissanayake A, Edwards J, Giraldo A, Hernandez-Restrepo M, Hyde KD, Jayawardena RS, Lombard L, Crous PW (2017) Genera of phytopathogenic fungi: GOPHY 1. Studies in Mycology 86(1): 99–216. https://doi.org/10.1016/j.simyco.2017.04.002

Miller MA, Pfeiffer W, Schwartz T (2010) Creating the CIPRES Science Gateway for inference of large phylogenetic trees. Proceeding of the 2010 gateway computing environments workshop (GCE), New Orleans, Louisiana, 1–8. https://doi.org/10.1109/GCE.2010.5676129

Quaedvlieg W, Binder M, Groenewald JZ, Summerell BA, Carnegie AJ, Burgess TI, Crous PW (2014) Introducing the consolidated species concept to resolve species in the Teratosphaeriaceae. Persoonia 33(1): 1–40. https://doi.org/10.3767/003158514X681981

Rambaut A (2009) FigTree, a graphical viewer of phylogenetic trees.

Ronquist F, Teslenko M, Van Der Mark P, Ayres DL, Darling A, Hohna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61(3): 539–542. https://doi.org/10.1093/sysbio/sys029

Sandoval-Denis M, Gene J, Sutton DA, Wiederhold NP, Cano-Lira JF, Guarro J (2016) New species of *Cladosporium* associated with human and animal infections. Persoonia 36(1): 281–298. https://doi.org/10.3767/003158516X691951

Schubert K, Groenewald JZ, Braun U, Dijksterhuis J, Starink M, Hill CF, Zalar P, de Hoog GS, Crous PW (2007) Biodiversity in the C*ladosporium herbarum* complex (Davidiellaceae, Capnodiales), with standardisation of methods for *Cladosporium* taxonomy and diagnostics. Studies in Mycology 58: 105–156. https://doi.org/10.3114/sim.2007.58.05

Stamatakis A (2006) RAxML-VI-HPC: Maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics 22(21): 2688–2690. https://doi.org/10.1093/bioinformatics/btl446

Swofford DL (2003) PAUP*: Phylogenetic analysis using parsimony (and other methods), version 4.0 b10. Sinauer Associates. https://doi.org/10.1111/j.0014-3820.2002.tb00191.x

Taylor JW, Jacobson DJ, Kroken S, Kasuga T, Geiser DM, Hibbett DS, Fisher MC (2000) Phylogenetic species recognition and species concepts in fungi. Fungal Genetics and Biology 31(1): 21–32. https://doi.org/10.1006/fgbi.2000.1228

Temperini CV, Pardo AG, Pose GN (2018) Diversity of airborne *Cladosporium* species isolated from agricultural environments of northern Argentinean Patagonia: Molecular characterization and plant pathogenicity. Aerobiologia 34(2): 227–239. https://doi.org/10.1007/s10453-018-9509-7

Vaidya G, Lohman DJ, Meier R (2011) SequenceMatrix: Concatenation software for the fast assembly of multi-gene datasets with character set and codon information. Cladistics 27(2): 171–180. https://doi.org/10.1111/j.1096-0031.2010.00329.x

White T, Bruns T, Lee S, Taylor JW (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis M, Gelfand D, Shinsky J, White T (Eds) PCR protocols: a guide to methods and applications 18(1): 315–322. https://doi.org/10.1016/B978-0-12-372180-8.50042-1

Zhang LB (2016) Eucommiaceae. Springer International Publishing, 193–196. https://doi.org/10.1007/978-3-319-28534-4_15

Zhang Y, Liu F, Wu W, Cai L (2015) A phylogenetic assessment and taxonomic revision of the thermotolerant hyphomycete genera *Acrophialophora* and *Taifanglania*. Mycologia 107(4): 768–779. https://doi.org/10.3852/14-173

Zhang ZF, Liu F, Zhou X, Liu XZ, Liu SJ, Cai L (2017) Culturable mycobiota from Karst caves in China, with descriptions of 20 new species. Persoonia 39(1): 1–31. https://doi.org/10.3767/persoonia.2017.39.01